Synthesis of a Benzomacrolactone-Based Somatostatin Mimetic

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The benzomacrolactone is a framework found in numerous natural products. The synthesis of an orthogonally functionalized benzomacrolactone from p-glucosamine and a salicylic acid derivative is described. This macrolactone was used for the synthesis of a somatostatin mimetic that has submicromolar affinity for the human somatostatin receptor 4 (hSSTR4).

A recent analysis of scaffolds investigated in organic chemistry has shown that a small number of frameworks are found in a large number of all known compounds.¹ This could, at least partially, explain why the diversity of scaffolds in a set of known drugs is considered to be low.² Hence the exploitation of frameworks less used to date in medicinal chemistry might ultimately lead to increased success in drug

(2) Bemis, G. W.; Murcko, M. A. J. Med. Chem. 1996, 39, 2887.

discovery. If the scaffolds are "natural product like", they could have increased biological importance.³

The low bioavailability and poor pharmacokinetics of somatostatin 1 has led to interest in nonpeptide mimetics which may be better drug candidates. $\overline{4}$ During the early 1990s, Hirschmann and colleagues at the University of Pennsylvania introduced the use of β -D-glucose, its enantiomers, and a diastereomer $(\beta$ -D-mannose) as scaffolds for the synthesis of ligands for the somatostatin, the substance

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⁽¹⁾ Lipkus, A. H.; Yuan, Q.; Lucas, K. A.; Funk, S. A.; Bartelt,W. F.; Schenck, R. J.; Trippe, A. J. J. Org. Chem. 2008, 73, 4443.

^{(3) (}a) Ganesan, A. Combinatorial Synthetic Design: The Balance of Novelty and Familiarity. In Exploiting Chemical Diversity for Drug Discovery; Bartlett, P. A., Entzeroth, M., Eds.; RSC Biomolecular Sciences, Royal Society of Chemistry: Cambridge, UK, 2006; pp 91-111. (b) Koch, M. A.; Waldmann, H. Drug Discovery Today 2005, 10, 471.

⁽⁴⁾ Hirschmann, R. Angew. Chem., Int. Ed. Engl. 1991, 30, 1278.

⁽⁵⁾ Hirschmann, R.; Nicolaou, K. C.; Pietranico, S.; Salvino, J.; Leahy, E. M.; Sprengeler, P. A.; Furst, G.; Smith, A. B., III; Strader, C. D.; Cascieri, M. A.; Candelore, M. R.; Donaldson, C.; Vale, W.; Maechler, L. J. Am. Chem. Soc. 1992, 114, 9217.

^{(6) (}a) Velter, I.; La Ferla, B.; Nicotra, F. J. Carbohydr. Chem. 2006, 25, 97. (b) Meutermans, W.; Le, G. T.; Becker, B. ChemMedChem 2006, 1, 1164. (c) Murphy, P. V.; Dunne, J. L. Curr. Org. Synth. 2006, 3, 403 and references cited therein.

P, and the β_2 -adrenergic receptors.⁵ Since this contribution, monosaccharides, 6 due to their polyfunctional nature, have been a basis for generating a wide range of compounds for screening. The glucopyranoside 3 (Scheme 1) prepared by the Hirschmann group, and containing the lysine, tryptophan, and phenylalanine side chains of 1, was found to be a ligand for somatostatin receptors, indicating that truncated mimetics are of interest. Other research has also indicated that interactions of the side chains of the Trp-Lys fragment of the hormone somatostatin 1 seem to be important for the recognition of SSTRs.⁷

Scheme 1

Herein we were interested in presenting groups on a benzomacrolactone scaffold that could mimic the interactions of the Trp-Lys fragment and selected 2 and analogues of 2 as targets for synthesis. There is no crystal structure available of somatostatin receptors binding to any ligands, but we hypothesized that the scaffold in 2 may span a larger surface of a protein receptor compared with a monosaccharide, which could lead to a ligand of higher affinity. Benzomacrolactones are common in nature (e.g., zearalenone, 8 Scheme 1), having a range of interesting biological properties,⁹ and analogues would be of interest for screening. Herein we describe the synthesis of somatostatin mimetics from a suitably decorated benzomacrolactone 4.

The preparation of 4 was envisaged (Scheme 1) from salicylic acid 5 and the alcohol 6. The incorporation of azide, OTIPS, and OPMB ($PMB = 4$ -methoxybenzyl) was based on providing the potential to introduce modifications selectively at these sites, with previous experience indicating orthogonality between the groups.¹⁰

Scheme 2

The synthesis of 6 (Scheme 2) commenced from D-glucosamine 7. The fully acetylated 2-azido-2-deoxy-D-glucopyranose was obtained as a mixture of anomers (α :β, 2:1) after diazo transfer and subsequent per-*O*acetylation.¹¹ A thioglycoside was then prepared,¹² and subsequent de-O-acetylation followed by reaction with dimethylanisaldehyde acetal and CSA gave 8.¹³ A TIPS protecting group was then introduced, and subsequent partial reductive cleavage gave 9. Next, 9 was hydrolyzed.¹⁴ If too high a proportion of water was used, it led to decomposition. The hemiacetal intermediate on reaction with N aBH₄ in EtOH/H₂O then gave 10.¹⁵ The 1, 2-diol of 10 was protected as a benzylidene acetal, and a pivalyl group was then introduced at the free alcohol group; subsequent partial reductive cleavage of the benzylidene acetal gave 11. The allylation of the free alcohol was carried out using sodium hydride and allyl iodide. The pivalyl ester¹⁶ was removed to give 6.

(14) Gomez, A. M.; Company, M. D.; Agocs, A.; Uriel, C.; Valverde, S.; Lopez, J. C. Carbohydr. Res. 2005, 340, 1872.

(15) Villalobos, A.; Danishefsky, S. J. J. Org. Chem. 1990, 55, 2776. (16) Nagorny, P.; Fasching, B.; Li, X. C.; Chen, G.; Aussedat, B.; Danishefsky, S. J. J. Am. Chem. Soc. 2009, 131, 5792.

(17) Rountree, J. S. S.; Murphy, P. V. Org. Lett. 2009, 11, 871.

^{(7) (}a) Angeles, A. R.; Neagu, I.; Birzin, E. T.; Thornton, E. R.; Smith, A. B., III; Hirschmann, R. Org. Lett. 2005, 7, 1121. (b) Chagnault, V.; Lalot, J.; Murphy, P. V. ChemMedChem 2008, 3, 1071.

⁽⁸⁾ Stob, M.; Baldwin, R. S.; Tuite, J.; Andrews, F. N.; Gillette, K. G. Nature 1962, 196, 1318.

⁽⁹⁾ For a review of resorcylic acid lactones see: Wissinger, N.; Barluenga, S. Chem. Commun. 2007, 22.

⁽¹⁰⁾ Murphy, P. V.; O'Brien, J. L.; Gorey-Feret, L. J.; Smith, A. B., III. Tetrahedron 2003, 59, 2259.

⁽¹¹⁾ Yan, R. B.; Yang, F.; Wu, Y. F.; Zhang, L. H.; Ye, X. S. Tetrahedron Lett. 2005, 46, 8993.

⁽¹²⁾ Yan, L.; Kahne, D. J. Am. Chem. Soc. 1996, 118, 9239.

⁽¹³⁾ Karst, N. A.; Islam, T. F.; Avci, F. Y.; Linhardt, R. J. Tetrahedron Lett. 2004, 45, 6433.

The preparation of the aromatic precursor 5 was next carried out (Scheme 3)^{17,18} from commercially available 12. Formation of the acetonide was first carried out, the resulting phenol was converted to its triflate, and a subsequent palladium-catalyzed coupling¹⁹ with potassium vinyltrifluoroborate gave 13. The acetonide 13 was then hydrolyzed to give 5. Reaction of 5 with the sorbitol derivative 6 by a Mitsunobu esterification²⁰ promoted by triphenylphosphine in the presence of DIAD gave 14. It is worth noting that the yield from the esterification decreased dramatically (from 83 to 45%) if the coupling was carried out starting from the benzyl ether 15. The pK_a of 5 is lower than that of 15 due to intramolecular H-bonding in 5, which explains the increased yield. The phenol was converted to its benzyl ether and then $RCM²¹$ using the Hoveyda-Grubbs II catalyst in $CH₂Cl₂$ under argon gave 4 (85% from 14). When 14 was directly subjected to RCM using the catalyst, the yield of the ring-closed product was low (∼40%). The optimum conditions for RCM were established after investigation of a variety of conditions, $22,23$ and the Hoveyda–Grubbs II catalyst²⁴ was chosen. Separation of the catalyst from the product was difficult. Chromatography using toluene/ $CH₃CN$ (200:1) as a solvent led to separation of the catalyst and isolation of 4 (92%).

To obtain somatostatin mimetics, side chains containing an amino group and indole group were installed on the scaffold (Scheme 4). First, the azide of 4 was reduced using $PMe₃,²⁵$ which was more effective than $PBu₃$ or $PPh₃$. The resulting amine was then coupled with indol-3-ylacetic acid 16^{26} to give 17. Next, the TIPS group was removed using TBAF in THF, and the resulting hydroxyl group alkylated using 1,4-dibromo-trans-2-butene 18 in the presence of Ag_2O^{27} gave 19. A variety of attempts to alkylate the secondary hydroxyl group using 20/21 failed. With the bromide 19 in hand, an amine surrogate then needed to be introduced. The Gabriel reagent (potassium phthalimide) was avoided as strongly basic conditions would have been possibly needed for removal of the phthalyl

Scheme 3

group, and there was evidence of decomposition under such conditions. Various N-benzyl derivatives were prepared, but these approaches were ultimately unproductive, and finally, the use of potassium $bis(t-butoxy$ $carbonyl$)amine (Boc₂NK) succeeded; its reaction with 19 giving 22 (77%) .²⁸

Scheme 4

- (18) Matos, M. C.; Murphy, P. V. J. Org. Chem. 2007, 72, 1803.
- (19) Molander, G. A.; Rivero, M. R. Org. Lett. 2002, 4, 107.
- (20) Kalivretenos, A.; Stille, J. K.; Hegedus, L. S. J. Org. Chem. 1991, 56, 2883.
- (21) Fuerstner, A.; Thiel, O. R.; Kindler, N.; Bartkowska, B. J. Org. Chem. 2000, 65, 7990.
- (22) Del Valle, J. R.; Goodman, M. J. Org. Chem. 2004, 69, 8946. (23) Hong, S. H.; Sanders, D. P.; Lee, C. W.; Grubbs, R. H. J. Am.
- Chem. Soc. 2005, 127, 17160.
- (24) Kingsbury, J. S.; Harrity, J. P. A.; Bonitatebus, P. J.; Hoveyda, A. H. J. Am. Chem. Soc. 1999, 121, 791.
- (25) Nyffeler, P. T.; Liang, C. H.; Koeller, K. M.; Wong, C. H. J. Am. Chem. Soc. 2002, 124, 10773.
- (26) Knor, S.; Khrenov, A. V.; Laufer, B.; Saenko, E. L.; Hauser, C. A. E.; Kessler, H. J. Med. Chem. 2007, 50, 4329.

Deprotection of 22 combined with reduction of the alkene groups was next investigated. In order to minimize the number of deprotection steps, trifluoroacetic acid (TFA) was first chosen as it could potentially remove the benzyl group from the phenol, as well as the Boc and PMB protecting groups in one pot.²⁹ The reaction of 22 with TFA in the presence of cation scavengers (PhSH, anisole, thioanisole) was screened,³⁰ and it was found that the use of thioanisole as a cosolvent with TFA led to the formation of 24 (60%), an analogue of 2. The reduction of the alkene groups of 22 was next investigated. Hydrogenation (5 or 10% Pd–C with AcOH or HCl in MeOH; 5% Pd(OH)₂, AcOH-MeOH; Wilkinson's catalyst, EtOH, etc.) of 22 led to decomposition. Moreover, it was found for some reagents that reduction of the indole ring 31 of 22 (as evidenced by MS analysis) was a problem, possibly due to the presence of a Boc group on the indole nitrogen. Hence a variety of conditions were screened $(H₂O, heat;³² Cs₂CO₃)$ imidazole, CH₃CN;³³ MeONa, MeOH) in order to remove the Boc group. Finally, the indole Boc and a second Boc group from the aminobutylene were removed from 22 using Zémplen conditions³⁴ at 40 °C to provide 23. Subsequent attempts at hydrogenation reactions with 23 were still problematic when $Pd - C$ or $Pd(OH)$ ₂ was used. Fortunately, hydrogenation from 23 using PtO₂ gave the desired intermediate, and subsequent treatment with TFA and thioanisole gave the target mimetic 2.

Somatostatin regulates the endocrine system and affects neurotransmission and cell proliferation via interaction with G-protein-coupled receptors (SSTRs). A sample of 2 $(< 95\%$ purity) showed activity as a ligand for hSSTR4

(27) Ley, S. V.; Tackett, M. N.; Maddess, M. L.; Anderson, J. C.; Brennan, P. E.; Cappi, M. W.; Heer, J. P.; Helgen, C.; Kori, M.; Kouklovsky, C.; Marsden, S. P.; Norman, J.; Osborn, D. P.; Palomero, M. A.; Pavey, J. B. J.; Pinel, C.; Robinson, L. A.; Schnaubelt, J.; Scott, J. S.; Spilling, C. D.; Watanabe, H.; Wesson, K. E.; Willis, M. C. Chem.⁻Eur. J. 2009, 15, 2874.

(29) Fletcher, S.; Gunning, P. T. Tetrahedron Lett. 2008, 49, 4817. (30) Masui, Y.; Chino, N.; Sakakibara, S. Bull. Chem. Soc. Jpn. 1980, 53, 464.

- (31) Shou, X. H.; Miledi, R.; Chamberlin, A. R. Bioorg. Med. Chem. Lett. 2005, 15, 3942.
	- (32) Wang, J.; Liang, Y. L.; Qu, J. Chem. Commun. 2009, 7601.
	- (33) Mohapatra, D. K.; Durugkar, K. A. ARKIVOC 2005, 20.
- (34) Ravinder, K.; Reddy, A. V.; Mahesh, K. C.; Narasimhulu, M.; Venkateswarlu, Y. Synth. Commun. 2007, 37, 281.

 $(K_i \sim 1 \,\mu\text{M})$. Compound 24 (>95% purity) was a ligand for hSSTR4 ($K_i = 0.58 \mu M$) and hSSTR5 ($K_i = 1.1 \mu M$). As the benchmark, 1 had K_i values of 0.52 nM (hSSTR4) and 0.75 nM (hSSTR5), whereas the mannosamine derivative 26

Figure 1. Somatostatin mimetics and affinities for hSSTR4.

(Figure 1, synthesis not shown) was inactive $(K_i > 100 \,\mu\text{M})$ against these receptor subtypes. Previous work from the Hirschmann group showed that 3 is a ligand for hSSTR4 with a $K_i = 1.1 \mu M$,³⁵ and work from our own group showed that 25 has a $K_i = 3.3 \mu M$ against this receptor.^{7b}

In summary, somatostatin mimetics based on benzomacrolactone scaffolds have been prepared. The approach could be extended to other peptidomimetics or other compounds for screening and broadens the potential for benzomacrolactone as a framework in bioorganic and medicinal chemistry.

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Supporting Information Available. NMR spectra, selected experimental procedures and analytical data for new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

⁽²⁸⁾ Allan, R. D.; Johnston, G. A. R.; Kazlauskas, R.; Tran, H. W. J. Chem. Soc., Perkin Trans. 1 1983, 2983.

⁽³⁵⁾ Hirschmann, R.; Hynes, J., Jr.; Cichy-Knight, M. A.; van Rijn, R. D.; Sprengeler, P. A.; Spoors, P. G.; Shakespeare, W. C.; Pietranico-Cole, S.; Barbosa, J.; Liu, J.; Yao, W.; Rohrer, S.; Smith, A. B., III. J. Med. Chem. 1998, 41, 1382.